

TECHNICAL DATA SHEET

Ornithine Decarboxylase Broth

Principle

Ornithine Decarboxylase Broth is modification of Moeller decarboxylase broth (1954-55). The media is composed of l-ornithine monohydrochloride, yeast extract, glucose and bromocresol purple. yeast extract provides nitrogen, vitamins, minerals, amino acids and growth factors. Glucose is carbon or energy source for the growth of microorganisms. L-ornithine monohydrochloride decarboxylated to form putrescine. Bromo cresol purple and cresol red are the pH indicators. When glucose is fermented by bacteria, the pH is lowered due to acid production, which changes the colour of the indicator from purple to yellow. Acid produced stimulates decarboxylase enzyme. Decarboxylation of ornithine yields putrescine in alkaline conditions, revert to purple colour. If the organisms do not produce decarboxylase enzyme, the colour of the medium remains yellow. glucose non-utilizers will not show any change in the medium color. Use light inoculate and do not read the tests after 24 hours incubation, as some organisms require longer incubation time of up to 4 days.

Use: For detection of the ability of microorganisms to decarboxylate ornithine.

Contents*

Ingredients	Gram/Litre
L-ornithine monohydrochloride	5.000
Yeast extract	3.000
Glucose	1.000
Bromocresol purple	0.015
pH at 25°C	6.8 ±0.2

***Formula adjusted for optimum performance and parameters**

Directions: Dissolved 9.00 grams in 1000 ml distilled water. Boil to dissolve the medium completely. Distribute 5.00 ml into the screw capped test tubes. Sterilize by autoclaving 15 lbs pressure (121°C) for 15 minutes. Allow to cool the tubes in an upright position and overlay with 2-3 ml of sterile mineral oil.

Specimens types analyzed

Water and food products etc.

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Precautions to be taken

All the handling, experiments, storage, and discarding should be performed with the help of trained and experienced technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving.

Performance and Evaluation

The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

Quality Control

Appearance	Light beige to greenish yellow color free flowing, homogeneous powder
Reaction of 0.9 % solution	6.8 $\pm$ 0.2 at 25 °C
pH	6.60-7.00
Color and clarity of ready medium	Purple color, clear slightly opalescent solution
Growth Promotion properties	Best at $\leq$ 100 CFU at 32-37 °C for 18-72 h
Indicative properties	Optimum at $\leq$ 100 CFU at 32-37 °C for 18-48 h
Negative control	Performed using sterile distilled water

Different Microbial Response: Cultural characteristics observed after an incubation at 35 $\pm$ 2°C for 18-24 hours. (Inoculated tubes are overlayed with sterile mineral oil).

Organism	ATCC	Inoculum	Color of medium
<i>Salmonella enterica</i>	9150	50-100	Purple color, positive reaction
<i>Salmonella typhimurium</i>	14028	50-100	Purple color, positive reaction
<i>Klebsiella aerogenes</i>	13048	50-100	Purple color, positive reaction
<i>Shigella flexneri</i>	9199	50-100	Yellow color, negative reaction
<i>Proteus mirabilis</i>	12453	50-100	Yellow color, negative reaction
<i>Escherichia coli</i>	25922	50-100	Variable reaction

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Storage and Shelf Life: The product is highly hygroscopic; keep the container tightly closed at all times and store it properly as per the conditions mentioned on the label. The declared expiry is valid only when stored as per the conditions mentioned on the label. Note: Sterilize media immediately after reconstitution.

Disposal: To avoid the contamination or propagation of any hazardous microbes used, unusable or modified preparation of this product must be disposed after autoclaving or incineration after completion of task.

Reference

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3. Downes F. P. and Ito K., (Ed.), (2001), Compendium of Methods for the Microbiological Examination of Foods, 4th Ed., American Public Health Association, Washington, D.C.
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