

## TECHNICAL DATA SHEET

### Phenol Red Mannitol Broth

#### Principle

Phenol red mannitol base is used for to study carbohydrate fermentation reaction. It is composed of proteose peptone, meat extract, sodium chloride, mannitol and phenol red. Peptone and meat extract provide carbon, nitrogen sources and essential growth factors required for growth of microorganisms. The sodium chloride maintains osmotic equilibrium of the medium. Phenol red is pH indicator dye. The medium contains all the necessary nutrients required for the growth of microorganism. The mannitol is added into the medium to carbohydrate fermentation reaction. If the organism has ability to ferment mannitol, with the acid or acid and gas production. Due to the acid production the color of medium changes to yellow. The gas production is detected by gas trapped in Durham's tubes.

**Use:** For determining the ability of microorganisms to ferment mannitol.

#### Contents\*

Ingredients	Gram/Litre
Proteose peptone	10.000
Meat Extract#	1.000
Sodium Chloride	5.000
Mannitol	5.000
Phenol red	0.018
pH at 25°C	7.4 ±0.2

\* Formula adjusted for optimum performance and parameters

# Equivalent to Beef Extract

**Directions:** Dissolved 21.00 grams in 1000 ml distilled water. Boil to dissolve the medium completely and distribute aseptically in test tubes containing inverted Durham's tubes. Sterilize by autoclaving at 15 lbs. pressure (121°C) for 15 min, cool it to 42-45 °C. Aseptically add filter sterilized or autoclave sterilized carbohydrate solution to sterile basal medium. Mix well and inoculate test sample aseptically.

#### Specimens types analyzed

Pure culture of microorganisms etc.

# OXFORD LAB FINE CHEM LLP

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**Regd Office:** Unit no 12, 1st Floor,  
Neminath Industrial Estate No.6,  
Navghar, Vasai (East), Palghar - 410210.  
Maharashtra, INDIA.

**Tel:** +91 250 2390032 / 2390989 / 2390990  
**Email:** sales@oxfordlabchem.com /  
info@oxfordlabchem.com  
**Web:** www.oxfordlabchem.com



## Precautions to be taken

These microbial media are intended for the in-vitro use only. All the handling, experiments, storage, and discarding should be performed with the help of skilled and knowledgeable technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving. Please go through the MSDS of the media to avoid any accidents or in emergency.

## Performance and Evaluation

The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

## Quality Control

Appearance	Light Beige to pink color free flowing, homogeneous powder
Reaction of 1.6% solution	7.4 ±0.2 at 25 °C
pH	7.20-7.60
Color and clarity of ready medium	red color, clear slightly opalescent solution
Growth Promotion properties	Best at ≤ 100 CFU at 32-37 °C for 18-72 h
Indicative properties	Optimum at ≤ 100 CFU at 32-37 °C for 18-48 h
Negative control	Performed using sterile distilled water

**Different Microbial Response: Cultural characteristics observed after incubation at 33-37°C for 18-24 hours. (Inoculum 50-100 CFU)**

Organism	ATCC	Growth	Acid	Gas
<i>Salmonella typhimurium</i>	14028	Luxurious	Positive	Positive
<i>Escherichia coli</i>	8739	Luxurious	Positive	Positive
<i>Shigella flexneri</i>	9199	Luxurious	Positive	Negative
<i>Klebsiella aerogenes</i>	13048	Luxurious	Positive	Positive

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**Storage and Shelf Life:** The product is highly hygroscopic; keep the container tightly closed at all times and store it properly as per the conditions mentioned on the label. The declared expiry is valid only when stored as per the conditions mentioned on the label. Note: Sterilize media immediately after reconstitution.

**Disposal:** To avoid the contamination or propagation of any hazardous microbes the used, unusable or modified preparation of this product must be disposed after autoclaving after completion of task.

## Reference

1. American Public Health Association, (1978) Standard Methods for the Examination of Dairy Products, 14<sup>th</sup> Ed., Washington
2. D.C Difco Manual (1998). 11<sup>th</sup> Edition. Difco Laboratories., Division of Becton Dickinson and Company, Sparks, Maryland, USA.
3. Monk, J. D., R. S. Clavero, L. R. Beuchat, M. P. Doyle, and R. E. Brackett. (1994). Irradiation inactivation of *Listeria monocytogenes* and *Staphylococcus aureus* in low-and high-fat, frozen and refrigerated ground beef. *J. Food Prot.* 57:969-974.
4. Wehr H. M. and Frank J. H., (2004), Standard Methods for the Microbiological Examination of Dairy Products, 17<sup>th</sup> Ed., APHA Inc., Washington, D.C.
5. Vera, H.D. 1950. Relation of peptones and other culture media ingredients to accuracy of fermentation tests. *Am. J. Public Health*, 40:1267-1272.

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