

TECHNICAL DATA SHEET

DNase Test Agar Base w/o DNA

Principle

DNase Test Agar Base is composed of tryptone, soya peptone, sodium chloride and agar. The DNase test agar base is used to detect DNA hydrolyzing microorganism particularly for identification of pathogenic Staphylococci. With the help of toluidine blue, it is also used for the identification of non-pigmented Serratia species, that might be improperly identified as Enterobacter and Klebsiella species. The media is fortified with deoxyribose nucleic acid which is used by the organisms to make nucleic acids and to use as a source of nitrogen, phosphate and carbon. In addition, tryptone and soya peptone are source of nitrogen, while sodium chloride maintains osmotic equilibrium of the medium.

Use: For detection of deoxyribonuclease activity of microorganisms & identification of pathogenic Staphylococci.

Contents*

Ingredients	Gram/Litre
Tryptone	15.00
Soya Peptone	5.00
Sodium chloride	5.00
Agar	15.00
pH at 25°C	7.3 ±0.2

* Formula adjusted for optimum performance and parameters

Directions: Dissolve 42.00 grams in 1000 ml distilled water. Boil to dissolve the medium completely. Sterilize by autoclaving at 121 °C at 15 lbs for 15 minutes. Cool to room 40 to 45 °C and distribute aseptically in petri plates with gentle shaking for equal distribution. Ensure complete solidification and inoculate test sample aseptically.

The media can be fortified with 0.1g toluidine blue before sterilization for proper visualization. DNase activity results in the production of a bright pink reaction due to the metachromatic property of Toluidine blue. Some strains of *Staphylococcus* may be inhibited on DNase Test Agar by the Toluidine blue. Further confirmatory tests for identification should be carried out.

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Specimens' types analyzed

Clinical and non-clinical samples, food and dairy samples etc.

Precautions to be taken

These microbial media are intended for the in-vitro use only. All the handling, experiments, storage, and discarding should be performed with the help of skilled and knowledgeable technicians and as per the established guidelines. The material should be disposed only after proper sterilization by autoclaving. Please go through the MSDS of the media to avoid any accidents or in emergency.

Performance and Evaluation

The expected performance of the medium is liable to use as per the direction on the label when stored at optimum conditions and within expiry date.

Quality Control

Appearance	Light beige colored free flowing, homogeneous powder
Reaction of 4.00% solution (0.20 g of deoxyribose nucleic acid)	7.3 ±0.2 at 25 °C
pH	7.10- 7.50
Gelling	Firm comparable with 1.5% agar gel
Color and clarity of ready medium	Light yellow colored opalescent gel or light blue with toluidine blue colored to slight opalescent gel
Growth Promotion properties	Best at ≤ 100 CFU at 32-37 °C for 18-72 h
Indicative properties	Optimum at ≤ 100 CFU at 32-37 °C for 18-48 h
Negative control	Performed using sterile distilled water

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Different Microbial Response

Cultural characteristics observed with added Toluidine Blue after an incubation at 35 - 37°C for 18 – 24 hours.

Organism	ATCC	Inoculum	Growth	Colony color
<i>Staphylococcus aureus</i>	25923	50-100	Luxurious	Positive, change in color from blue to pink purple around the growth when toluidine blue is used/ clear zone surrounding colonies when plates are flooded w/1N HCL
<i>Staphylococcus epidermidis</i>	12228	50-100	Luxurious	Negative

Storage and Shelf Life: The product is highly hygroscopic; keep the container tightly closed at all times and store it properly as per the conditions mentioned on the label. The declared expiry is valid only when stored as per the conditions mentioned on the label. Note: Sterilize media immediately after reconstitution.

Disposal: To avoid the contamination or propagation of any hazardous microbes the used, unusable or modified preparation of this product must be disposed after autoclaving after completion of task.

Reference

1. American Public Health Association, Standard Methods for the Examination of Dairy Products, (1978), 14th Ed., Washington D.C.
2. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock, D.W. (2015). *Manual of Clinical Microbiology*, 11th Edition. Vol. 1.
3. Salfinger Y., and Tortorello M.L. Fifth (Ed.), (2015), *Compendium of Methods for the Microbiological Examination of Foods*, 5th Ed., American Public Health Association, Washington, D.C.
4. Wehr H. M. and Frank J. H., (2004), *Standard Methods for the Microbiological Examination of Dairy Products*, 17th Ed., APHA Inc., Washington, D.C.

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